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First report of the tick *Hyalomma scupense* (natural vector of bovine tropical theileriosis) on the French Mediterranean island of Corsica

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ABSTRACT

Hyalomma scupense (Acari, Ixodidae) is a common tick species found in several areas in North Africa, Asia and South Europe and an efficient natural vector of bovine tropical theileriosis (*Theileria annulata*), a livestock disease with an important economic impact. For one year, 1938 ticks were collected on cattle in several Corsican slaughterhouses; 168 of them were morphologically identified as *H. scupense*. This result was confirmed by genetic identification using sequences of mitochondrial cytochrome c oxidase subunit I (COI) and ribosomal internal transcribed spacer 2 (ITS2) genes. The presence of 2 different stages (adults and nymphs), collected in various areas of the island, indicates that a population of *H. scupense* is established in Corsica. However, bovine tropical theileriosis has not been diagnosed on the island so far.

KEY WORDS

Hyalomma scupense ; Cattle ; Molecular identification ; *Theileria annulata* ; Corsica

1 - INTRODUCTION

Hyalomma scupense occurs in a wide area ranging from northern Africa, Sudan, Turkey, the Middle East, southern Russia and central Asia, to China; it has also been reported from some southern European countries (Apanaskevich et al., 2010). Taxonomic uncertainty as to the identity of *Hyalomma* (*Euhyalomma*) *scupense* Schulze, 1919 and *Hyalomma detritum* Schulze, 1919 has existed for a long time, but it is now established that the two names correspond to the same species; *H. scupense* is the valid name and *H. detritum* is a synonym (Apanaskevich et al., 2010). It is the most abundant ixo-did tick infesting cattle in Morocco and Tunisia (Estrada-Pena et al., 2004; Gharbi and Darghouth, 2014). This tick species is monotropic and has become endophilic in many regions. *H. scupense* is a one- or two-host tick; domestic cattle are the most common hosts for adults and immature stages, but almost all ungulates may be infested. Single records are also known from other hosts (domestic dogs, hares (*Lepus* sp.), foxes (*Vulpes* sp.)) and humans are often attacked by the adults (Hoogstraal, 1956; Apanaskevich et al., 2010). It is one of the most important vectors of bovine tropical theileriosis (*Theileria annulata*). This protozoan is a major pathogen in the Maghreb region, causing high economic losses in cattle (milk yield decrease, weight loss, abortions, high treatment costs and deaths; Gharbi et al., 2011). *H. scupense* is also a vector of other diseases, including Q-fever (*Coxiella burnetii*) and equine piroplasmiasis (*Theileria equi*) (Hoogstraal, 1956). Although *H. scupense* can probably transmit the virus of Crimean–Congo hemorrhagic fever (CCHF), it does not appear to be a major vector. Hoogstraal (1979) quoted a study by Chumakov et al. (1973) in which CCHF virus infection was demonstrated by the fluorescent antibody test in the salivary glands of *H. scupense* in Kzyl-Orda Oblast, Kazakhstan. According to Estrada-Pena et al. (2012), *H. scupense* is one of the main *Hyalomma* species implicated in the transmission of the virus in Eurasia, but no other evidence of this is given.

Corsica is a French Mediterranean island situated 15 km north of Sardinia and 90 km west of Tuscany in Italy (Fig. 1). The island is 183 km long and 83 km wide; it is the most mountainous island in the Mediterranean area (with Monte Cinto peaking at 2706 m). Its climate is classified as hot summer Mediterranean. About 300,000 people live in Corsica and livestock farming is an important economic activity with approximately 150,000 sheep, 48,000 goats, 40,000 pigs and 70,000 cattle. Cattle farming is of a very extensive type, the animals (mainly local *Bos taurus* cattle) live outside all year and usually without any barn.

Neither the tick fauna of Corsica nor the transmitted pathogens have been fully investigated. However, a few ad hoc studies showed the existence of three species of the genus *Hyalomma* on the island. (i) *Hyalomma marginatum* is well-established, widespread, and found on many hosts (Morel, 1959; Pérez-Eid, 2007; personal observations). It is one of the main vectors of the zoonotic CCHF virus and an experimental vector of *Theileria annulata* but unlikely to play a significant role in the field (Estrada-Pena et al., 2004). It is also a vector of *Rickettsia* spp., including *Rickettsia aeschlimanni* in Corsica (Matsumoto et al., 2004), and of *Babesia caballi*, the agent of equine babesiosis (Estrada-Pena et al., 2004). According to Hoogstraal (1956) it is also a natural reservoir of the agent of Q-fever (*C. burnetii*); (ii) *Hyalomma aegyptium* is mainly found on tortoises although it occasionally infests other hosts, small mammals (hedgehogs, hares, rodents, etc.), lizards and even birds (partridges). It has been identified only once in Corsica (one male) on a tortoise *Testudo hermanni* (Matsumoto et al., 2004); (iii)

Hyalomma rufipes, a tropical sub-Saharan species, has been collected on migrating birds and may play a role in the dissemination of *R. aeschlimanni*, with which it has been found infected in Corsica (Matsumoto et al., 2004). This species is known to be the most important vector of CCHF virus in southern Africa and it also transmits *Anaplasma marginale* and *Babesia occultans* to cattle (Estrada-Pena et al., 2004). Nymphs of this species are commonly found on birds migrating from Africa to Europe (Hoogstraal et al., 1963) and isolated adults have even been reported in Northern Europe.

A better knowledge of the Corsican tick fauna is needed to determine present and potential dangers to human and animal health related to tick-borne pathogens. This was the aim of a systematic tick survey carried out in 2014–2015. The finding of the species *H. scupense* during this study is presented here in advance because of its great potential importance as an efficient vector of *T. annulata* on cattle.

2 - MATERIALS AND METHODS

From May 2014 to May 2015, ticks were collected in the 3 Corsican cattle slaughterhouses (Ponte-Leccia, Cuttoli and Porto-Vecchio, Fig. 1), from cattle living in different areas of the island. The national cattle identification system (ear tags) allowed to trace back the origin of the cattle. The slaughterhouses of Ponte-Leccia and Cuttoli were visited monthly, whereas samples were collected only three times in Porto-Vecchio. During each visit, the whole skins of 15–25 cows were examined, just after flaying, and the ticks were manually collected. They were stored in 70% ethanol at -20°C until their identification according to their morphological characteristics using a stereomicroscope, with the keys and descriptions by Hoogstraal (1956), Walker et al. (2003), Estrada-Pena et al. (2004) and Apanaskevich et al. (2010).

To confirm the morphological identification of *H. scupense*, genomic DNA (gDNA) was extracted and purified from either one or two legs, using the QIAamp DNA microkit following manufacturer protocol (Qiagen). The use of only one or two legs allowed both morphological and molecular identification and the remainder of the specimen was kept for examining the presence of pathogenic organisms. Target genomic regions included the mitochondrial cytochrome c oxidase subunit I (COI) genes, as well as the whole sequences of the ribosomal internal transcribed spacer-2 (ITS2) region. The primers used for COI amplification were LCO1490 (5'-GGTCAACAAATCATAAAGATATTGG-3') and HCO2198 (5'-TAAACTTCAGGGTGACCAAAAATCA-3') (Folmer et al., 1994). The primers used for ITS2 were H1 (5'-TGGCTTCGTCTGTCTGAGGGTC-3') and H2 (5'-TGCTTAAATTCAGCGGGTTGTC-3'). Conventional polymerase chain reaction (PCR) was performed in a total reaction volume of 25 μL consisting of 2 μL of gDNA, 1X Colourless GoTaqflexi buffer (Promega, Madison, WI, USA), 1.5 mM MgCl_2 (Promega, Madison, WI, USA), 0.2 mM of each dNTP (Promega, Madison, WI, USA, reference U1420), 0.4 μM forward primer, 0.4 μM reverse primer and 1U GoTaq G2 hotstart Polymerase (Promega, Madison, WI, USA, reference M7405). DNA amplification consisted in an activation step of 95°C for 4 min, followed by 35 cycles of 95°C for 40 s, 52°C for 40 s, and 72°C for 1 min (COI) or 1 min 40 (ITS2), and a final extension at 72°C for 5 min. PCR products were visualized on Labchip GX1 (PerkinElmer). Positive products were sent to Genoscreen (France) for sequencing in both directions using dilutions of the original PCR primers. The sequences were then assembled and manually edited using Sequencher (GeneCodesCorp) to remove any ambiguities between

strands and sequences. Genetic distance between sequences obtained and reference sequences from GenBank were calculated using Mega V6.0

3 - RESULTS

A total of 1,938 ticks were collected from cattle, among which 168 were identified as *H. scupense* (70 females, 74 males and 24 nymphs), representing 8.7% of the collected ticks (Fig. 2). Ticks were obtained from 72 “communes” (smallest administrative unit in France, Fig. 1) and *H. scupense* were collected from cattle raised in 21 of them. It represented more than 50% of the collected ticks in 10 communes. Individuals from this species were collected from December to May, with a peak in January–March, during which *H. scupense* constituted 40–53% of the ticks infesting the slaughtered cattle. The 24 nymphs were collected from December to March

Eleven specimens morphologically identified as *H. scupense* (4 males, 4 females and 3 nymphs) were submitted to molecular identification. Ten COI and 8 ITS2 sequences were obtained. Sequence divergence was found within the COI sequences (4 haplotypes; GenBank accession numbers, AN: KT598361, KT598360, KT598362, KT598363). The genetic distance within Corsican specimens was 0.002 (standard deviation of 0.001). No sequence variation was found within ITS sequences (AN: KT598364). All sequences confirm the morphological identification of *H. scupense*. Table 1 shows mean pairwise distance of all the sequences obtained for COI, for ITS2 and reference sequences. The genetic distance between the Corsican specimens and *H. scupense* was 0.7% for both genes while the minimum genetic distance between the Corsican specimens and the other species was 7.7%.

4 - DISCUSSION

In spite of the large geographic distribution of *H. scupense* in the world, its presence in Corsica and its relative abundance (8.7% of the collected ticks) were not expected since the species was rarely found in the areas neighboring the island where significant tick studies have been carried out. Two specimens were recorded in Sardinia, on deer (Garippa et al., 2003) and on man (Montarsiet al., 2011); and 7 questing adults (designated as *H. detritum*) were collected on the vegetation on the Tuscan island of Pianosa (Tomassone et al., 2013). It is not frequent in continental Spain (Oteo et al., 2006) and has not been found on Minorca Island (Ros-García et al., 2012). In continental France, the tick was reported in the South-West where it seems to be established (Macaigne and Pérez-Eid, 1993) though the collected numbers were low. But in Northern Africa where tropical theileriosis is a major cattle disease, *H. scupense* is often the most widespread tick species on cattle, representing between 84.3 and 99.1% of the ticks population in Tunisia (in farms affected by tropical theileriosis), 30.8% in Morocco (Gharbi and Darghouth, 2014) and 13.6% in Algeria (Benchikh Elfegounet et al., 2013). In Corsica, *H. scupense* was reported in almost 30% of the investigated areas (Fig. 1) and two different stages (adults and nymphs) were found several times on the same animals. These observations provide strong arguments for the existence of an endemic and well-established population of *H. scupense* ticks in Corsica, which is rather surprising for an area of south-western Europe.

The way *H. scupense* arrived in Corsica remains unexplained. The role played in the dispersion of ticks by migratory birds has to be considered, although this species was so far not found on migrating birds (e.g., Hoogstraal et al., 1963; Wallménius et al., 2014). Cattle are the main hosts of *H. scupense* and although there is currently no introduction of animals from countries where the tick is established to Corsica, this could have happened earlier. As the Corsican tick fauna had never been fully investigated before this study, it cannot be excluded that *H. scupense* may be a long-time established species. A phylogenetic analysis of several *H. scupense* populations, from Corsica but also from Maghreb countries, could perhaps give clues regarding the origin of Corsican ticks.

Although its seasonal dynamics have not been systematically studied as yet, it appears that the habitat and life cycle of *H. scupense* are different in Corsica and in the Maghreb countries, for two reasons. *H. scupense* is usually described, in the latter regions, as an endophilic tick (Estrada-Pena et al., 2004; Gharbi and Darghouth, 2014) with infestations often associated with barns, stables, sheds and pens; because cattle are kept outside in Corsica, the tick can not be endophilic. Secondly, maximum activity of the species was observed from January to March in this study, which is quite different from what Gharbi and Darghouth (2014) described in North Africa with a peak adult activity from May to August.

Theileria buffeli/orientalis, the causative agent of benign theileriosis, has been reported in Corsica (Uilenberg, 2000) but bovine tropical theileriosis (*T. annulata*) has not been diagnosed so far. *T. annulata* is only transstadially transmitted, so the immature *H. scupense* stages cannot be infected before feeding on an infested host. If indeed the Corsican population of *H. scupense* is presently not infected, migrating birds coming from enzootic areas would not constitute a risk of introducing *T. annulata*. Such birds have so far not been found infested by immature *H. scupense*, and moreover, as larvae (and therefore the resulting nymphs) are never infected, the greatest potential danger of *T. annulata* introduction would be the importation of live carrier cattle from an infected country.

The occurrence of *H. scupense* in Corsica highlights however the risk of transmission of *T. annulata* to the local cattle population. Bovine tropical theileriosis occurs in many neighboring Mediterranean countries, but has not been reported in continental Italy and Sardinia. It is present in Sicily (Loria et al., 1999), the Spanish Island of Minorca (Ros-García et al., 2012) and continental Spain (García-Sanmartín et al., 2006). In Sicily and continental Spain the main vector of *T. annulata* is *Hyalomma lusitanicum* while the vector species is uncertain in Minorca. In the other countries of the Mediterranean area, the main vector appears to be *H. scupense*. Using partial amplification and sequencing of the 18S rRNA gene, Criado-Fornelio et al. (2003) reported the identification of *T. annulata* in the blood of a bovine animal reared in France. Also Bonnet et al. (2013) detected *T. annulata* DNA in a single *Dermacentor marginatus* collected in the field by flagging in southern France (Aveyron). This tick species is not known as a vector of *T. annulata* and finding the pathogen in the tick does not necessarily mean that the tick can transmit it. These two reports certainly need further confirmations and if tropical theileriosis occurred in southern France, it would probably have been detected. Because of the extensive livestock management and the rustic local cattle breed, the infection might however remain undetected in Corsica. Bovine tropical theileriosis is one of the most economically important diseases and for the future of eventual improvements on the animal husbandry in Corsica, it is of the utmost importance to determine whether it is present or not on the island.

CONFLICT OF INTEREST

The authors declare no conflicts of interest.

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REFERENCES

- Apanaskevich, D.A., Filippova, N.A., Horak, I.G., 2010. The genus *Hyalomma* Koch, 1844. X. Redescription of all parasitic stages of *H. (Euhyalomma) scupense* Schulze, (= *H. detritum* Schulze) (Acari: Ixodidae) and notes on its biology. *Folia Parasitol.* 57, 69–78.
- Benchikh Elfegoun, M.-C., Gharbi, M., Djebir, S., Kohil, K., 2013. Dynamique d'activité saisonnière des tiques ixodidés parasites des bovins dans deux étages bioclimatiques du nord-est algérien. *Revue Elev. Méd. Vét. Pays Trop.* 66, 117–122.
- Bonnet, S., de la Fuente, J., Nicolle, P., Liu, X., Madani, N., Blanchard, B., Maingourd, C., Alongi, A., Torina, A., Fernández de Mera, G., Vicente, J., George, J.-C., Vayssier-Taussat, M., Joncour, G., 2013. Prevalence of tick-borne pathogens in adult *Dermacentor* ticks from nine collection sites in France. *Vector-Borne Zoonotic Dis.* 13, 226–236.
- Chumakov, M.P., Smirnova, S.E., Shalunova, N.Y., Mart'yanova, L.I., Fler, G.N., Zgurskaya, G.N., Maksimov, S.S., Kasymov, K.T., Pak, T.P., 1973. Proofs of ecological identity of Crimean hemorrhagic fever and Central Asian hemorrhagic fever. *Abstr. Inv. Pap.* 9. *Int. Congr. Trop. Med. Malar.* (Athens), 33–54.
- Criado-Fornelio, A., Martínez-Marcos, A., Buling-Sarana, A., Barba-Carretero, J.C., 2003. Molecular studies on *Babesia*, *Theileria* and *Hepatozoon* in southern Europe. Part I. Epizootiological aspects. *Vet. Parasitol.* 113, 189–201.
- Estrada-Pena, A., Bouattour, A., Camicas, J.L., Walker, A.R., 2004. *Ticks of Domestic Animals in the Mediterranean Region: A Guide to Identification of Species*. University of Zaragoza, Zaragoza.
- Estrada-Pena, A., Jameson, L., Medlock, J., Vatansever, Z., Tishkova, F., 2012. Unraveling the ecological complexities of tick-associated Crimean–Congo Hemorrhagic Fever virus transmission: a gap analysis for the Western Palearctic. *Vector-Borne Zoonotic Dis.* 12, 743–752.
- Foata, J., Mouillot, D., Culioli, J.L., Marchand, B., 2006. Influence of seasonal hostage on wild boar parasites in Corsica using indicator species analysis. *J. Helminthol.* 80, 41–45.
- Folmer, O., Black, M., Hoeh, W., Lutz, R., Vrijenhoek, R., 1994. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Mol. Mar. Biol. Biotech.* 3, 294–299.
- García-Sanmartín, J., Nagore, D., García-Pérez, A.L., Juste, R.A., Hurtado, A., 2006. Molecular diagnosis of *Theileria* and *Babesia* species infecting cattle in Northern Spain using reverse line blot macroarrays. *BMC Vet. Res.* 2, 16–22.
- Garippa, G., Biddau, M., Cherchi, M., 2003. Endo ed ectoparassiti di *Cervus elaphus corsicanus* della Sardegna. *J. Mt. Ecol.* 7 (Suppl), 189–190.
- Gharbi, M., Darghouth, M.A., 2014. A review of *Hyalomma scupense* (Acari, Ixodidae) in the Maghreb region: from biology to control. *Parasite* 21, 2.
- Gharbi, M., Touay, A., Khayeche, M., Laarif, J., Jedidi, M., Sassi, L., Dhargouth, M.A., 2011. Ranking control options for tropical *theileriosis* in at-risk dairy cattle in Tunisia: using benefit-cost analysis. *Rev. Sci. Tech. Off. Int. Epiz.* 30, 763–778.
- Hoogstraal, H., 1956. *African Ixodoidea. Volume I. Ticks of the Sudan*. Research Report NM, Cairo, Egypt, 1101 pp.
- Hoogstraal, H., 1979. The epidemiology of tick-borne Crimean–Congo hemorrhagic fever in Asia, Europe and Africa. *J. Med. Entomol.* 15, 307–417.

Hoogstraal, H., Kaiser, M.N., Traylor, M.A., Gaber, S., Guindy, E., 1963. Ticks (Ixodoidea) on birds migrating from Africa to Europe and Asia 1959–61. *Bull. World Health Organ.* 24, 197–212.

Loria, G.R., Riili, S., Vitale, F., Greco, A., Sparagano, O., 1999. Clinical and laboratory studies on *theileriosis* outbreaks in Sicily, Italy. *Parassitologia* 41 (Suppl. 1), 63–67.

Macaigne, F., Pérez-Eid, C., 1993. *Hyalomma scupense* Schulze 1919 (Acarina, Ixodoidea) tique autochtone du sud-ouest de la France. *Ann. Parasitol. Hum. Comp.* 68, 199–200.

Matsumoto, K., Parola, P., Brouqui, P., Raoult, D., 2004. *Rickettsia aeschlimanni* in *Hyalomma* ticks from Corsica. *Eur. J. Clin. Microbiol. Infect. Dis.* 23, 732–734.

Montarsi, F., Maioli, G., Fois, F., Mereu Piras, P., Magliano, A., Pascucci, I., Toma, L., Mignone, W., Torina, A., Capelli, G., 2011. Uncommon ticks from Italy. In: *Proceeding of the 7th International Conference Ticks and Tick-borne Pathogens (TTP7)*, Zaragoza, p. 213.

Morel, P.-C., 1959. Les *Hyalomma* (Acariens: Ixodidae) de France. *Ann. Parasitol. Hum. Comp.* 34, 552–555.

Oteo, J.-A., Portillo, A., Santibañez, S., Perez-Martinez, L., Blanco, J.-R., Jimenez, S., Ibarra, V., Pérez-Palacios, A., Sanz, M., 2006. Prevalence of spotted fever group *Rickettsia* species detected in ticks in La Rioja, Spain. *Ann. N. Y. Acad. Sci.* 1078, 320–323.

Pérez-Eid, C., 2007. Les tiques. Identification, biologie, importance médicale et vétérinaire. Lavoisier, Editions TEC&DOC, 2007, 314 pp.

Ros-García, A., García-Pérez, A.L., Verdera, J., Juste, R.A., Hurtado, A., 2012. Monitoring piroplasm infection in three cattle farms in Minorca (Balearic Islands, Spain) with previous history of clinical piroplasmosis. *Vet. Parasitol.* 190, 318–325.

Tomassone, L., Grego, E., Auricchio, D., Iori, A., Gianni, F., Rambozzi, 2013. Lyme borreliosis spirochetes and spotted fever group rickettsiae in ixodid ticks from Pianosa Island, Tuscany Archipelago, Italy. *Vector-Borne Zoonotic. Dis.* 13, 84–91.

Uilenberg, G., 2000. Tick-borne infections of cattle on Corsica. *ICTTD newsletter n°14*, 2000, 13 (ISSN: 1385-3422).

Walker, A.R., Bouattour, A., Camicas, J.-L., Estrada-Pena, A., Horak, I.G., Latif, A.A., Pegram, R.G., Preston, P.M., 2003. *Ticks of Domestic Animals in Africa. A Guide to Identification of Species.* Bioscience Reports, Edinburgh, Scotland, UK221.

Wallménius, K., Barboutis, C., Fransson, T., Jaenson, T.G., Lindgren, P.-E., Nyström, F., Olsen, B., Salaneck, E., Nilsson, K., 2014. Spotted fever *Rickettsia* species in *Hyalomma* and *Ixodes* ticks infesting migratory birds in the European Mediterranean area. *Parasit. Vectors* 7, 318.

FIGURE 1

Localization of *Hyalomma scupense* and proportion of this species among the ticks collected on cattle in Corsica

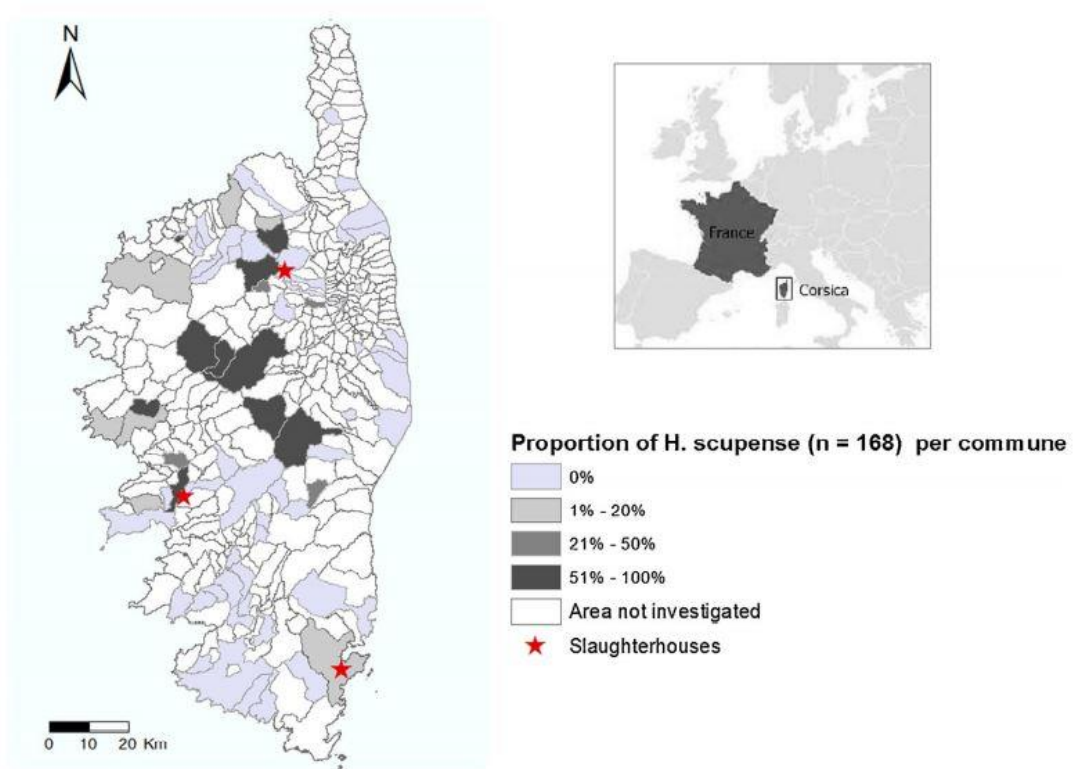


FIGURE 2

Dorsal view of a male of Corsican *Hyalomma scupense*



TABLE 1

Pair-wise genetic distances between COI and ITS2 of reference sequences and Corsican specimens (accession numbers are between parentheses)

COI		1	2	3	4	5	6
1.	<i>H. dromedarii</i> (AJ437083)						
2.	<i>H. anaticum</i> (KF912622.2)	8.7%					
3.	<i>H. asiaticum</i> (KF527440.1)	14.7%	16.9%				
4.	<i>H. lusitanicum</i> (EU827743)	9.6%	9.3%	15.5%			
5.	<i>H. marginatum</i> (EU827693.1)	4.0%	8.7%	14.6%	8.2%		
6.	<i>H. scupense</i> (KM235712.1)	12.6%	15.2%	9.8%	13.6%	11.7%	
7.	Corsican specimens (n = 10)	13.3%	15.3%	10.9%	14.7%	13.3%	0.7%
ITS2		1	2	3	4		
1.	<i>H. asiaticum</i> (JX845148)						
2.	<i>H. anaticum</i> (HQ005303)		11.1%				
3.	<i>H. marginatum</i> (JQ737104)		11.1%	4.6%			
4.	<i>H. scupense</i> (JX845150)		8.6%	8.3%	8.3%		
5.	Corsican specimens (n = 8)		7.8%	7.6%	7.7%		0.7%